Seeing is Believing: Enabling Functional Imaging of NanoLuc[®] Technologies with Bioluminescence Microscopy

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1. Introduction

Research for new therapeutics often use cell-based assays with microplate readers to generate relative light unit (RLU) and relative fluorescence unit (RFU) values for cellular responses. However, it is becoming increasingly desirable for researchers to want to view their cell models to correlate image results with traditional microplate reader results. This is particularly important for assays that measure protein translocation, internalization, degradation of target proteins and cellular organelles, as well as cellular processes like cell migration, cell proliferation and apoptosis.

Using imaging technologies, researchers can validate appropriate expression and localization, identify rare cellular events, differentiate responders versus non-responders, perform outlier removal, and analyze mixed cell populations. Today, fluorescence microscopy is widely used as a tool to accomplish these tasks. However, the use of fluorescence comes with several limitations, including 1) limited sensitivity, 2) high background, 3) fluorophore photo-toxicity, and 4) photo-bleaching.

Here we introduce an affordable, benchtop microscope capable of luminescence, fluorescence and brightfield microscopy to enable imaging of Nano luciferase-based assays, including NanoLuc[®], NanoBRET[™] and NanoBiT[®].

2. GloMax[®] Galaxy Bioluminescence Imager

- Study protein dynamics and cellular physiology
- Living & fixed cells & tissues
- Use NanoLuc[®] technologies for rare events, assay validation, analysis of mixed cell populations



Entry-level, benchtop system



- gas)

3. Imaging Low Abundance Endogenous Proteins

Binary Complementation of NanoBiT® Enzyme (High affinity) Relative protein expression in various CRISPR HeLa lines 1.5×10 1×10⁷ CASP3 CFL EGFR HDAC2 HDAC6 **CRISPR HeLa Clonal Lines**



Figure 2. To assess the capability of GloMax[®] Galaxy to resolve luminescence from low abundant endogenous proteins, a HiBiT / LgBiT complementation assay was used (Panel A). Endogenous protein targets were HiBiT-tagged using CRISPR Cas9 in HeLa cells. LgBiT was then expressed ectopically in each cell model. Luminescence from each target was measured with the GloMax[®] Discover Microplate Reader (Panel B) to determine RLU signals to compare with bioluminescence images taken from GloMax[®] Galaxy and constitute a 1.5 log range in expression. Bioluminescence images were captured using 1 minute exposure times for Cofilin (Panel C) EGFR (Panel D) and HDAC2 (Panel E). HDAC6 (Panel F) and CASP3 (Panel G) represented very low expressing proteins and they were exposed for 3 minutes (Panel F) and 5 minutes (Panel G).

Figure 5. GloMax[®] Galaxy control and acquisition software interface.

7. Interior View and Accessories



Figure 6. GloMax® Galaxy interior view

Figure 6. GloMax[®] Galaxy fluorescence excitation modules (Panel A). They System will be supplied with 1 swappable fluorescence excitation module; additional modules for sale as accessories. The System will be supplied with a filter slide containing 3 fluorescence excitation filters and 1 open position. Al 4 emission filter positions can be customized. A stage top incubator and gas controller for long-term kinetic imaging (Panel B) will be available as an accessory time.

8. System Specifications

Simple to use

 Designed for experienced and n quickly collect publication-quality

Multiple imaging modalities

 Complement the power of Nano fluorescence capabilities.

Affordable

Accessible to all types of labora

Live cell kinetics

 Control gas, temperature and hu study biology in real time.

Complete solution provider

 Technical support for the instruit and experiment.



Available late 2024.

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	SPECIFICATIONS	
ovice users to ty images.	Dimension (W x H x D)	14.7 in x 18.8 in x 21.0 in (37.3cm x 47.7cm x 53.3cm)
	Weight	62 lb (28 kg)
oLuc [®] with	Sample Vessels	Slide, microchamber, 35mm dish, 6, 12, 24, 48, and 96-well plates
	Capture Modes	Brightfield, Fluorescence, Luminescence, and Filtered Luminescence
atories.	Excitation Source	LED, transillumination
umidity to	Objective	Nikon 20X Plan APO Lambda D, 0.75 NA, 1mm WD
	System Magnification	10.4X
ment, assays	Sensor and Pixel Size	CMOS, 3200x2200 pixels, 4.5μm x 4.5μm pixel size
	Maximum Field of View	1.4mm x 0.95mm
	Resolution Limit	1.3 to 2.0μm
	Environment Control	Optional: Stagetop chamber and Controller with built-in gas mixer

Expand your research capabilities and use of NanoLuc[®] technologies with an affordable bioluminescence imaging system.

