

Dehydrated Human Amnion Chorion Membranes Contain Key Extracellular Matrix Proteins,

Resist Rapid In Vitro Degradation, and Function as a Scaffold

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INTRODUCTION

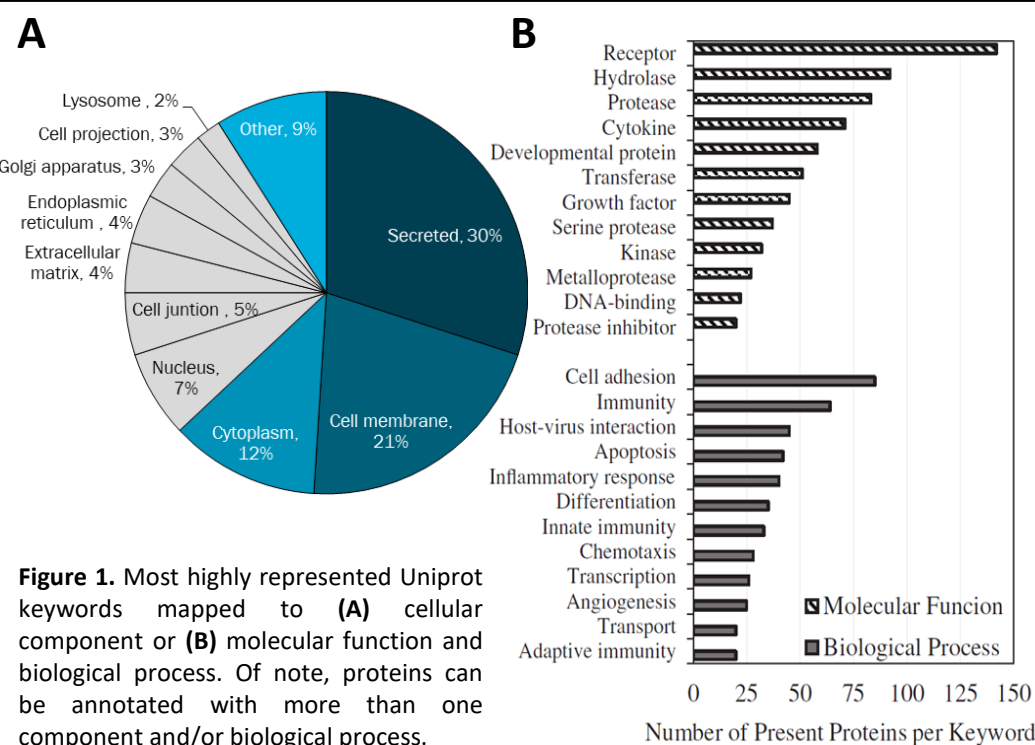
Placental-based products have a long-documented use in various wound applications. They have been shown to contain growth factors and cytokines and an extracellular matrix (ECM) rich in collagens, fibronectin, and other ECM molecules. Here, a minimally manipulated dehydrated amnion/chorion membrane (dACM[®]) was evaluated for biophysical structure and composition as well as its ability to function as a scaffold using *in vitro* models.

[®]NuShield[®], Organogenesis, Canton, MA

METHODS

- Preservation of key regulatory proteins was assessed using a high-throughput multiplex array, while biophysical structure was assessed using immunohistochemistry (IHC).
- Quantification of key extracellular matrix proteins was evaluated using colorimetric assays.
- Reduction of proteolytic activity was evaluated using a fluorescent cleavage assay.
- Degradation characteristics, including mass loss and collagen content, were assessed using an *in vitro* simulated wound fluid (SWF) degradation model for up to 10 days.
- Scaffold functionality was evaluated using a fibroblast cell attachment and proliferation model on intact (non-degraded) and 3-day SWF-degraded dACM.

HIGH-THROUGHPUT PROTEOMIC EVALUATION



BIOPHYSICAL STRUCTURE AND CHARACTERIZATION OF dACM

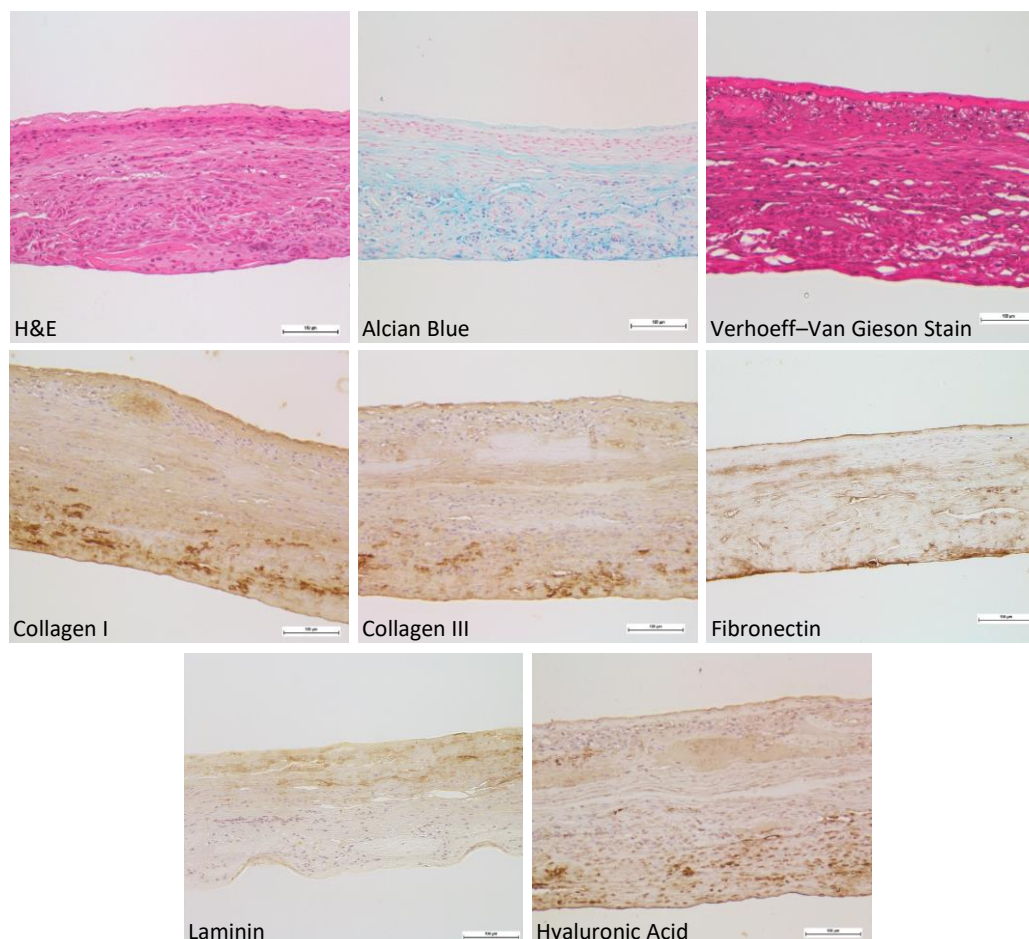


Figure 2. Biophysical structure of dACM. Histological evaluation of dACM including H&E, Alcian Blue, and Verhoeff-Van Gieson staining. Immunohistochemistry of key extracellular matrix proteins maintained in dACM. Collagens I and III and fibronectin were highly concentrated in the chorion layer (lower portion of grafts), while laminin was predominately found in the basement membrane of the amnion. Scale bars = 100 μ m.

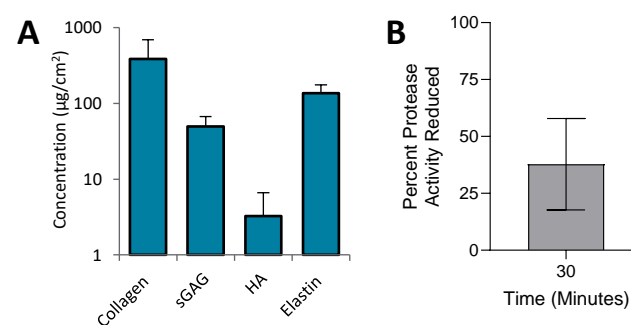


Figure 3. Characterization of dACM. (A) dACM maintains key extracellular matrix proteins, with collagens and elastin being present in the highest concentrations. (B) Reduction of protease activity with dACM resulted in ~40% reduction compared to uninhibited controls. Average \pm stdev reported.

PHYSICAL SCAFFOLD PROPERTIES

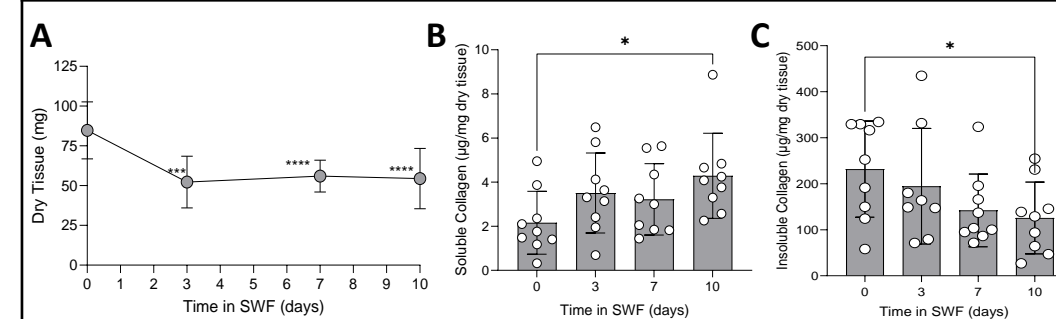


Figure 4. Physical scaffold properties after exposure to SWF in an *in vitro* model. (A) Dry tissue weight and (B) soluble and (C) insoluble collagen content were evaluated over 10 days. Average \pm stdev reported. * $P \leq 0.05$, *** $P \leq 0.001$, **** $P \leq 0.0001$ compared to T0.

dACM SUPPORTS CELL ATTACHMENT FOLLOWING DEGRADATION

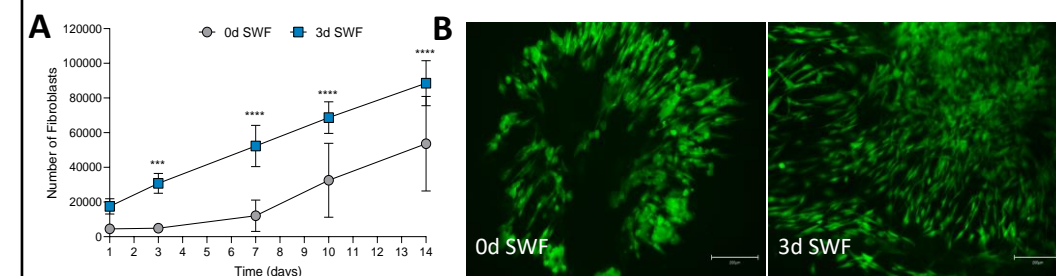


Figure 5. Cellular scaffold properties after exposure to SWF. (A) Fibroblast attachment and growth over 14 days on intact (0d) and 3-day (3d) SWF-degraded dACM and (B) representative Calcein AM-stained images of live fibroblasts after 3 days of attachment (20x; 200 μ m scale bar). Average \pm stdev reported. *** $P \leq 0.001$, **** $P \leq 0.0001$ compared to T0.

CONCLUSIONS

- Structurally, dACM maintained 640 regulatory proteins that were mapped to a variety of molecular functions and biological processes.
- The distribution and localization of key extracellular matrix proteins reveals that collagens I and III, in addition to elastin, fibronectin, laminin, hyaluronic acid, and glycosaminoglycans, are found throughout dACM.
- dACM reduced protease activity, with approximately 40% of activity diminished after 30 minutes.
- Exposure to SWF demonstrated that dACM is resistant to rapid degradation, as significant mass loss was only observed from T0 to day 3.
- dACM served as a scaffold for fibroblast attachment and proliferation, and these properties were maintained through degradation, with significantly greater attachment seen on partially degraded matrices compared to intact.
- These results demonstrate the complex structure of dACM, along with its ability to function as a scaffold for fibroblast attachment and growth.